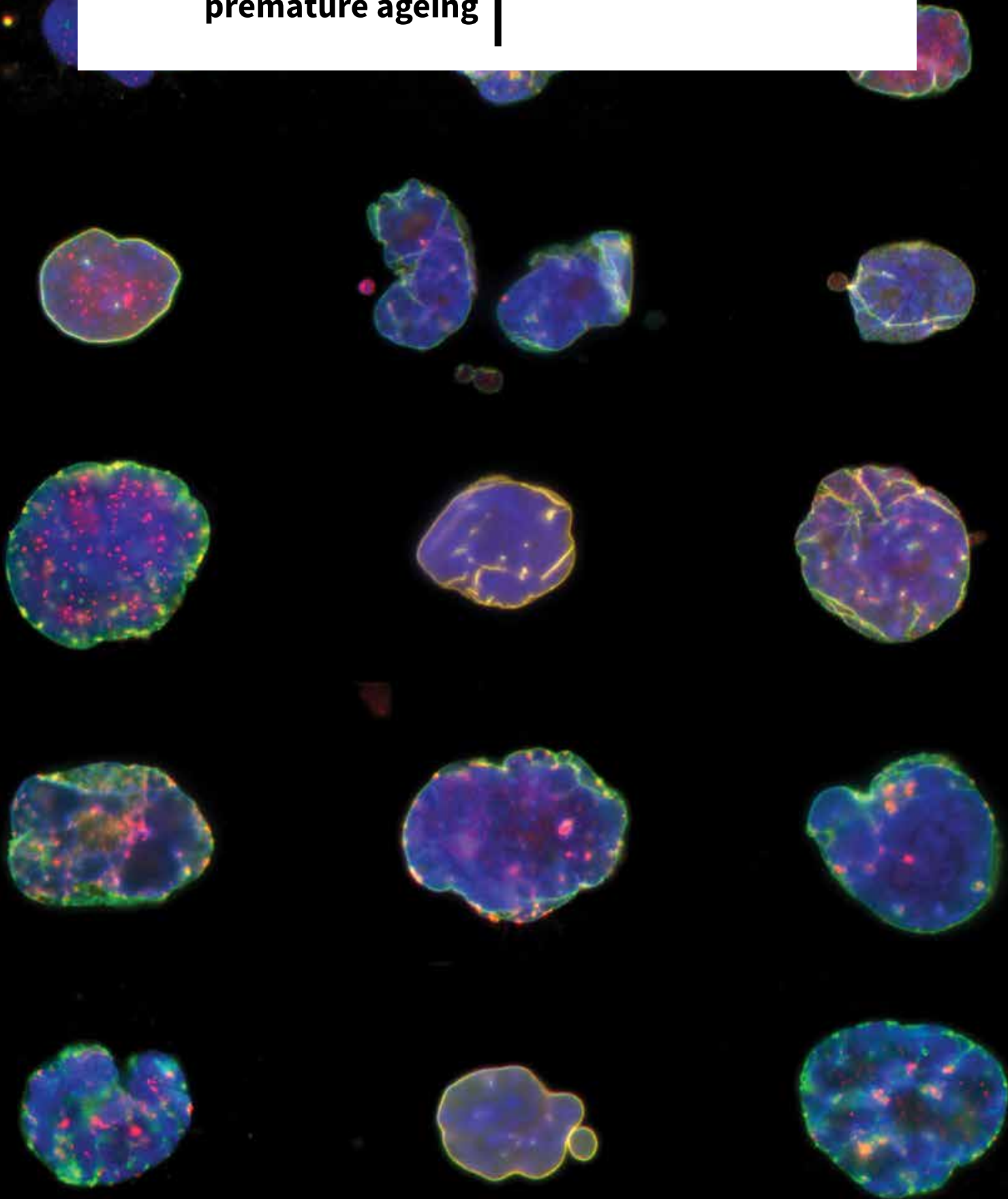
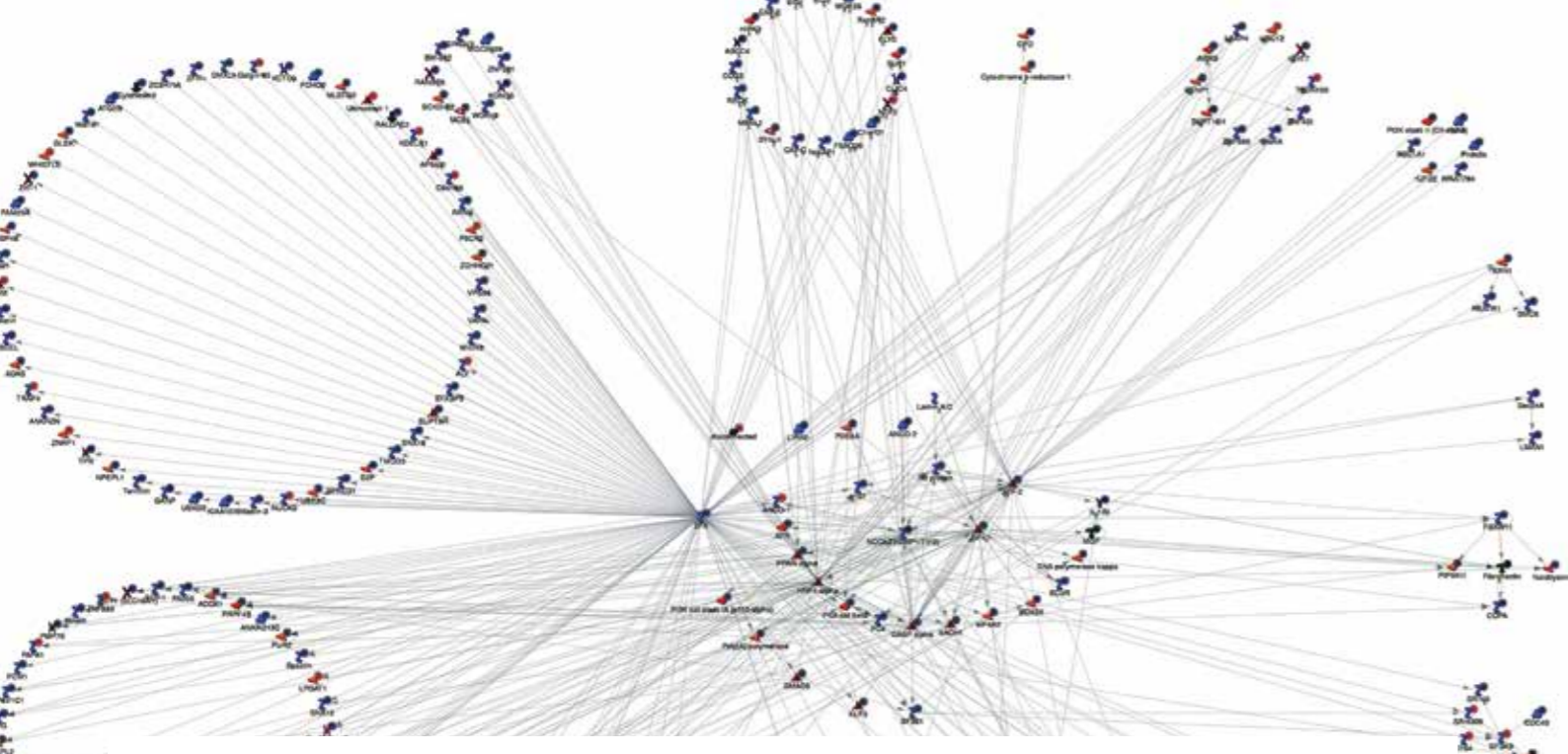


Putting a stop to premature ageing

Professor Karima Djabali



Hutchinson-Gilford Progeria Syndrome is caused by mutations on the LMNA gene which encodes lamin A and C proteins that are critical components of the nuclear envelope that protects the genome integrity.



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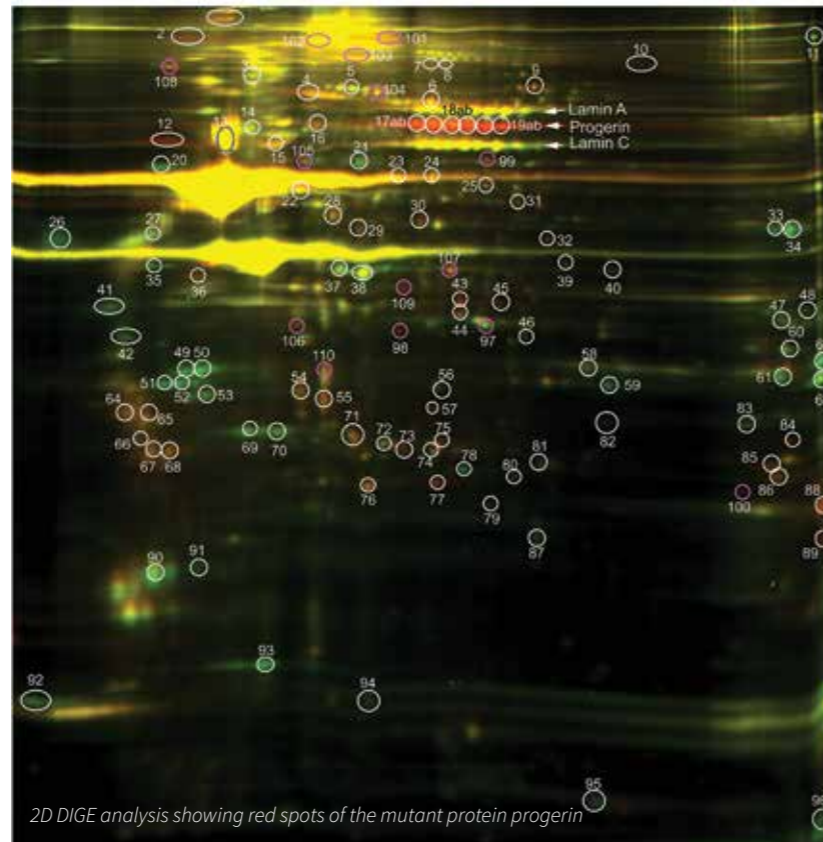
For over ten years, **Professor Karima Djabali** and her team have been on a mission to find a cure for Hutchinson-Gilford Progeria Syndrome. This cutting edge research can inform both interventions for this fatal disorder and reveal further insights into normal ageing.

What is premature ageing?

Hutchinson-Gilford Progeria Syndrome (HGPS) is a harrowing diagnosis to receive. This rare genetic disorder can be characterised by premature ageing, particularly of the vascular system, leading to severe atherosclerosis (narrowing of the arteries) and consequently, stroke or heart attack. Other symptoms of the disorder include growth delay, loss of body fat and osteoporosis. Young people diagnosed with HGPS have a life expectancy of just seven to 20 years. So what is being done to combat this tragic disease? Professor Djabali and her team are working to define the molecular and cellular processes that lead to the development of HGPS and to find effective therapeutic interventions for children affected by the condition.

Discovering the Mechanisms of Disease

There have been significant gains in knowledge surrounding HGPS pathophysiology since 2003. Researchers associated with the Progeria Research Foundation research focus now know that HGPS is caused by mutations on the LMNA gene, the role of which is to provide



instructions for the production of lamin A and lamin C. Lamins are proteins which line the inside of the nuclear envelope and play an important role in the organisation and the scaffolds of the nucleus of the cell. There are two categories of these proteins: A-type lamins and B-type lamins. In almost 90% of HGPS cases, mutation occurs on a part of the LMNA gene which leads to the deletion of 50 amino acids in the lamin A protein. The deletion of these amino acids means that the cells can no longer produce mature lamin A and instead a mutant form of lamin A (known as progerin) is formed. Progerin then accumulates in the nucleus and causes severe deformations in the nuclear membrane and major reorganisation of the chromatin. This affects the cell cycle and cell migration and promotes premature senescence (the deterioration of cellular function).

The cardiovascular system is the primary target of HGPS. Progerin accumulates predominantly in the nuclei of the vascular cells with the consequence of dramatically

accelerated cardiovascular disease in young patients with HGPS. The disease process of HGPS is similar to that of the vascular pathology of ageing – they both feature hardening and narrowing of the arteries and small blood vessels, as well as prominent adventitial fibrosis (the scarring and thickening of the outer layer of the arteries.)

Most recently, Professor Djabali and her team reported on how the build-up of progerin in the nucleus impairs chromosome maintenance during mitosis. Progerin accumulation disrupts cell division by displacing centromere protein F, a protein which attaches the chromosome to the spindle fibre in the middle phase of mitosis. The end result is delayed reformation of the nuclear envelope, genomic instability and cells with two nuclei rather than one. The cumulative effect of these defects with each round of mitosis then predisposes cells to premature dysfunction.

Journey to a cure

With each study, Professor Djabali and her colleagues get one step closer to a cure. Their 2010 paper explains the influence of protein farnesylation on the pathology of HGPS and how this could be inhibited. Protein farnesylation in this case refers to the retention of a farnesyl group (a carbon-based organic compound) to prelamin A, the precursor to lamin A. As stated above, the LMNA mutation associated with HGPS leads to the deletion of amino acids, including the amino acid which usually signals the cleavage site for the farnesyl group from prelamin A in order to form mature lamin A. Instead, the farnesyl group remains attached making progerin toxic to the nucleus. The team's research suggests that this impacts the lamin A-retinoblastoma protein (pRB) signalling network, causing pRB to interact differently with its downstream transcription factors and regulators, thus implicating this network as a key factor in HGPS pathogenesis. Now that the mechanism starts to unravel, the team treated HGPS

fibroblasts with a protein farnesyltransferase inhibitor (FTI) and found that this reversed abnormal gene expression in 288 of the 352 genes found in fibroblasts from HGPS patients. This outcome suggests that modulation of the lamin A-pRB signalling network could be a key factor in slowing down premature ageing.

A 2012 study by Professor Djabali and her colleagues provided the first in vivo evidence (evidence from within a living organism) that progerin is produced in adult stem cells (visualised on skin sections). Through the development of a new method of isolating naïve multipotent skin derived precursor cells from human fibroblast cultures, the team were able to observe a number of cell properties. These precursor cells had similar stem cell properties to cells isolated from skin biopsies. The cells could self-renew and differentiate in smooth muscle cells and fibroblasts. They were also found to express nestin (a marker of neuronal stem cells). A subset of these cells exhibited progerin positive signals which could reiterate the same molecular process which leads to progerin accumulation in adult stem cells. This study provides a viable alternative to skin biopsy samples from HGPS patients, which are not readily available for research, and opens new avenues to study the pathogenesis of other rare diseases.

In a 2014 study, Professor Djabali and her team investigated the use of sulforaphane (SFN) as a treatment for HGPS. SFN is an organic compound which can be found in cruciferous vegetables, such as young broccoli and cauliflower sprouts. The team aimed to test the ability of SFN to induce progerin clearance and improve disease phenotype in HGPS cell cultures using two-dimensional differential in a gel electrophoresis (2D-DIGE) approach. Progerin accumulation results in alterations to several components of protein degradation pathways. It leads to the impairment of autophagy (the cell's ability to degrade and recycle cellular components) and proteasome activity (the breakdown of damaged or unneeded proteins). As a potent inducer of antioxidant and detoxification enzymes, SFN also enhances the protein degradation systems and promotes progerin clearance and proteostasis (the process by which cells control the quantity and folding of proteins). SFN does this through modulation of the nuclear factor erythroid 2-related factor (Nrf2) signalling pathway.

The Nrf2 signalling pathway can be looked upon as a 'master regulator' of antioxidant defence within the cells. Nrf2 targets genes which promote resistance to cancer and other diseases through the transcription of detoxifying and anti-inflammatory proteins. It may also play a role in preventing premature ageing. So how does SFN fit in? It's all down to the interaction between SFN and a sensor known as Keap1. Keap1 is part of the body's stress response system and inhibits Nrf2 signalling. SFN disrupts this process and allows Nrf2 to accumulate in the nucleus and activate the transcription of its target genes some of which were identified as downregulated in HGPS cells in the 2D-DIGE analysis.

In the study, cells treated with SFN exhibited significant increases in growth rate, proteasome activity and autophagy. SFN treatment also led to more efficient DNA damage repair in the cell, increased ATP levels and normalisation of the nuclear envelope. Significantly, the increase in proteostasis and progerin clearance remained high over 12 weeks, supporting its efficacy as a long term therapeutic strategy in cell based system and show potential for children with HGPS. Therefore, Professor Djabali's team are conducting further studies to determine the extent to which modulation of the Nrf2 pathway with SFN could be a candidate target for future clinical trials.



HGPS patient, image courtesy of PRF

Additionally, stimulating the Nrf2 pathway may be beneficial to normal cells, progerin and prelamin A also accumulate in some cells of the skin and vascular system in unaffected individuals. Because the number of cells expressing this abnormal protein – progerin – seems to increase with age, further studies are underway to test the potential of manipulating this pathway in the context of physiological ageing. Remarkably, these studies show once again that important insight into the ageing process can be gained by studying HGPS. HGPS may not only pinpoint therapeutic avenues for fighting this rare genetic disorder, but might also teach us more about normal ageing and provide clues for how to slowdown ageing processes in general.

The Latest Cutting Edge Research

More recently, the findings of a 2016 study by Professor Djabali and her colleagues further explain the underlying mechanisms of genomic instability in HGPS cells. By investigating the effects of progerin on the dynamics of mitosis, the team uncovered evidence which appears to directly link progerin action to defects in chromosome segregation, nuclear envelope reformation and other cell defects. The first part of the study analysed the distribution of proteins in relation to the components of the nuclear lamina and envelope during mitosis. The findings showed that progerin began to cause defects in chromosome segregation as early as the metaphase (shortly after the breakdown of the nuclear envelope). This led to delays in nuclear envelope reformation as well as trapped cell components and proteins in the endoplasmic reticulum during cell division.

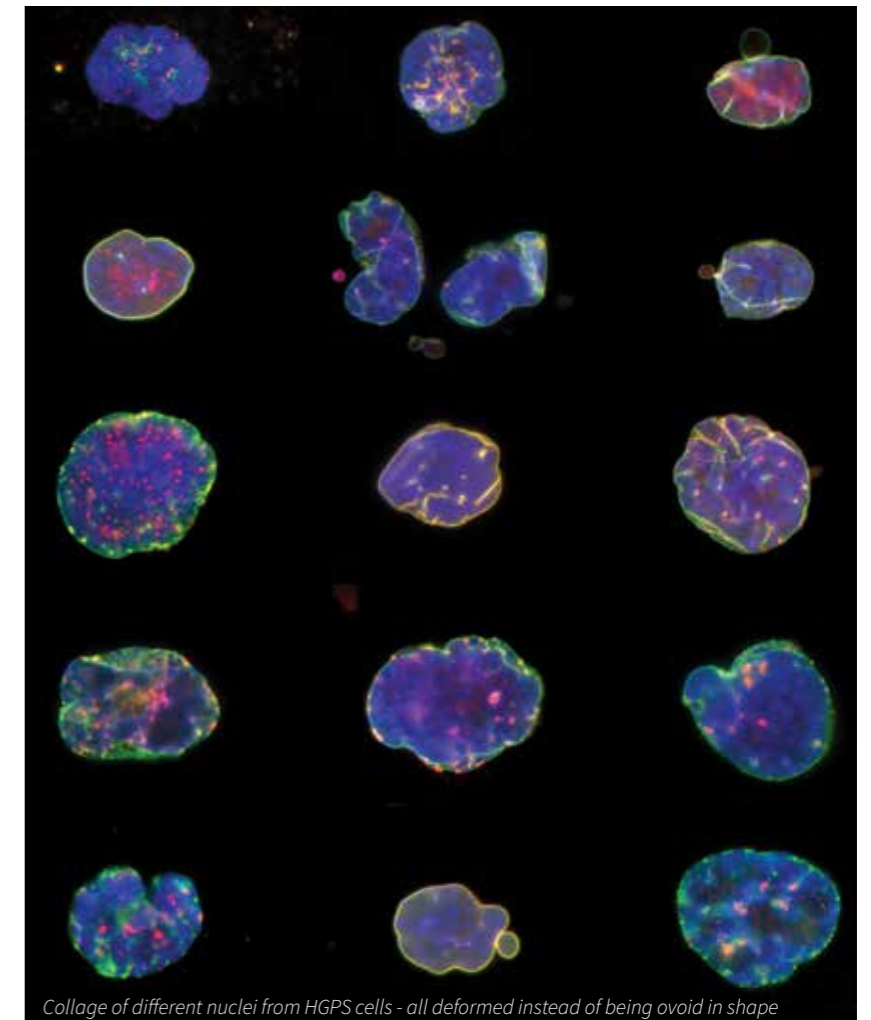
The second part of the investigation looked at the effect of progerin on the localisation of two proteins, CENP-E and CENP-F, on kinetochores.

Kinetochores are protein structures which allow chromatids to attach to spindle fibres and are required for proper segregation of chromosomes. While distribution of CENP-E was similar to that in normal cells, CENP-F was displaced from kinetochores during the metaphase. This suggests that CENP-F could be a target of progerin during mitosis. The team also found evidence of increased chromatin lagging and genomic instability which increased proportionally with progerin levels. These progerin dependent defects accumulate with each round of mitosis, suggesting that high levels of progerin predispose cells to undergo premature senescence.

Delaying Age Related Disease

There is also a lot to learn from this research regarding physiological ageing outside of disease processes. By examining the processes underlying premature ageing syndromes, Professor Djabali and her colleagues can determine whether or not similar mechanisms underlie normal ageing. It may also lead to insights in the pathology underlying other age related diseases, particularly vascular and neurological disorders affecting the elderly population. Therefore, the team aim to define isolate and characterise skin stem cells in order to determine how ageing might impact stem cell renewal and differentiation. This unique approach to the study of ageing makes the laboratory the only one in Germany to examine the characteristics of ageing from this angle.

So far, studies into HGPS have led to some interesting discoveries in the processes of ageing. An examination of skin biopsies from individuals across the life span showed a comparable frequency of progerin at a low level in all age groups, but increased levels and deeper distribution of the protein were found in the older population. This suggests that progerin expression could be used as a biomarker for normal cellular ageing. Likewise, in a histological comparison of vascular pathology in HGPS and vascular pathology of ageing, the team and collaborators detected progerin in both samples, supporting the theory that progerin has a role in cardiovascular ageing amongst the general population. Similar results were found in a later study examining the link between atherosclerosis in ancient humans, premature ageing syndromes and normal ageing. After imaging studies of ancient mummies revealed vascular



Collage of different nuclei from HGPS cells - all deformed instead of being ovoid in shape

calcification, researchers became interested in the possibility of a genetic predisposition to atherosclerosis. In their 2014 study, the team found that progerin can be detected in vascular cells. The resulting loss in cells from the vascular walls and replacement by fibrous tissues promotes hardening and calcification of the blood vessels. Although vascular progerin is found in low levels in the general population, these levels increase with age, suggesting a potential role for progerin in age-related atherosclerotic processes.

Next steps in ageing research

So what's next for Professor Djabali and her team? As well as continuing her search for a cure for HGPS, the knowledge gained from the research on progeria could drive future research in Alzheimer's disease (AD). By dissecting the underlying mechanisms of ageing cells in patients with AD, it may be possible to better understand which processes are inducing neuronal degeneration.

Research into treatments for HGPS may

also be useful in identifying new targets for cancer therapy. In simple terms, cancer is the uncontrolled division and growth of cells. One of the major risk factors for cancer is ageing. What is remarkably intriguing is that cellular ageing leads to cell cycle exit via down regulating signalling pathways controlling the proliferation of cells. In cancer some of the overlapping signalling triggers the opposite outcome. How does the balance between these signals dictate such opposite cellular fate? Can HGPS teach us which signalling or gene targets could lead to anti-cancer treatments? Professor Djabali is searching for answers.

Finally, a greater understanding of how adult stem cells work can inform work in regenerative medicine and drug therapies. Studying the mechanisms by which protein breakdown regulates the processes of ageing could be the key to identifying therapeutic interventions which promote healthy cellular function and ameliorate age-related pathologies.



Meet the researcher

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Professor Karima Djabali is a faculty Professor at the school of Medicine, Technical University Munich. She was previously Assistant Professor in the Department of Dermatology in Columbia University, New York, and scientist at the CNRS, France. Since receiving her PhD in Biology from the University of Paris 7, Professor Djabali has undertaken a number of prestigious postdoctoral fellowships and research positions and has received grants from the National Institute of Health, the Alexander von Humboldt Foundation, the Progeria Research Foundation, and the DFG. She has also received a number of awards for her research, including the Irving Clinical Research Career Award and Christine Kühne Center for Allergy Research and Education (CK-CARE) Award. For more than a decade, Professor Djabali and her team have focused their research on cellular ageing under normal and disease conditions, specifically in Hutchinson-Gilford progeria syndrome a very rare segmental premature aging syndrome. Her lab also aims to define, isolate and characterise adult stem cells for basic science, disease modeling and therapy. Together with outstanding international collaborators, she hopes that one day they can defeat HGPS disease and possibly slow down other age-related conditions.

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